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(54) Title: **TOTAL PROTEIN DETECTION METHODS AND DEVICES**

(57) Abstract: An assay for the determination of protein in an aqueous test fluid that comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye also has affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein to thereby render the assay suitable for the detection of total protein in the test fluid. The buffer and dye may be absorbed in a test strip of absorbent material.

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TOTAL PROTEIN DETECTION METHODS AND DEVICES

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FIELD OF THE INVENTION

The present invention relates generally to methods and devices of detecting proteins. Specifically, the invention is directed to methods and devices for detecting proteins at lower pH's using various buffers.

10

BACKGROUND OF THE INVENTION

Methods for the detection of urinary proteins are often more sensitive to albumin than to other urinary proteins. It is important, however, to detect proteins other than albumin, especially in the case of Bence-Jones proteinuria. The detection of proteins should cover a wide range of protein concentrations since the decision levels and recommended actions to be taken by clinicians will vary depending on the concentration of protein detected in a patient's urine. For example, persistent proteinuria greater than 50 mg/dL represents strong evidence of renal disease whereas a protein level of greater than 300 mg/dL is consistent with a diagnosis of nephrotic syndrome. A concentration of protein in urine of greater than 800 mg/dL suggests massive protein loss and warrants a renal biopsy and/or steroid therapy. Accordingly, it is apparent that a test for protein in urine should be effective over a wide range of protein values.

Various methods for the determination of protein in aqueous fluid have been reported in the literature. These methods include the Kjeldahl method, biuret method, Lowery method, dyestuff combination method, UV method and fluorometric method. In general, proteins react with various substance including dyes such as bromphenol blue, coomassie brilliant blue and eosine, as well as metal ions such as silver (I), copper (II), zinc (II) and lead (II).

Urine contains amounts of salt that vary between individuals. Some of these salts may act as a buffer when a diagnostic assay demands operation at a pH different than that of the urine sample. For example, urine sample with salts such as bicarbonate, acetate or phosphate tend to resist the lowering of pH. Creatinine, a common component of urine, may also act as a buffer in resisting the lowering of pH. The result of this buffering is that

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some urine samples will cause a strip pH to be higher than optimal. A higher than optimal strip pH may result in color generation of the protein indicator dyes, providing a false indication of the presence of protein. It is desired to have a buffering system that reduces such urine effects, while not interacting with the chemistry occurring.

Accordingly, a need exists for a method and device for detecting proteins over a wide range of clinical concentrations and pH's.

SUMMARY OF THE INVENTION

According to one embodiment, an assay for the determination of protein in an aqueous test fluid comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye has an affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein, to thereby render the assay suitable for the detection of total protein in the test fluid.

According to another embodiment, a dry device for use in determining protein levels in a fluid test sample comprises an absorbent material having absorbed therein a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye has an affinity for protein such that it will provide a detectable response in the presence of greater than 15 mg/dL protein upon contact between the device and the fluid test sample, to thereby render the device suitable for the detection of total protein in the fluid test sample.

According to a further embodiment, an assay for the determination of protein in an aqueous test fluid comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, or combinations thereof. The buffer is added in

sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range and which has an affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein, to thereby render the assay suitable for the detection of total protein in the test fluid.

According to yet a further embodiment, a dry device for use in determining protein levels in a fluid test sample comprises an absorbent material having absorbed therein a buffer and a dye. The buffer is selected from citrulline, malonic acid or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid within a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range and which has affinity for protein such that it will provide a detectable response in the presence of greater than 15 mg/dL protein upon contact between the device and the fluid test sample, to thereby render the device suitable for the detection of total protein in the fluid test sample.

DETAILED DESCRIPTION OF THE ILLUSTRATED EMBODIMENT

The assay for protein of the present invention can be carried out in either the wet or dry format. It is most conveniently carried out in the form of an absorbent test strip impregnated with (a) a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof, and (b) a dye having a pKa which enables it to operate as a protein indicator (e.g., a color indicator) at a pH of from about 2.0 to about 3.0.

Buffers

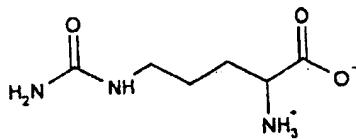
The buffer to be used in the present invention is added in sufficient quantity to maintain the pH of the assay including the test fluid within a range of from about 2.0 to about 3.0. The buffer may have a prior adjustment of pH that is commonly accomplished by adding acid or base to a solution of the buffer chemical to achieve a particular pH, or by adding a mixture of the buffer chemical and its salt in selected proportions to give a particular pH. The buffer is preferably added to maintain the pH of the assay including the test fluid within about +/- 0.2 pH units of the selected target pH. The buffer is

preferably added in sufficient quantity to maintain the pH of the assay including the test fluid within a range of from about 2.4 to about 3.0.

The preferred buffers are citrulline, malonic acid or combinations thereof. Buffers may be selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. Various concentrations of these buffers may be used, but the concentrations of the buffers are generally from about 200 to about 500 mM.

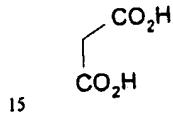
Citrulline may be in the form of L-citrulline, D-citrulline and D,L-citrulline and is represented by structure A.

10

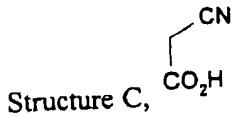


Structure A

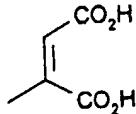
Malonic acid that is useful in the present invention is represented by Structure B,



Cyanoacetic acid that is useful in the present invention is represented by



Structure C, Citraconic acid that is useful in the present invention is represented by Structure D.



Structure D

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Methyl phosphonic acid that is useful in the present invention is represented by Structure E.

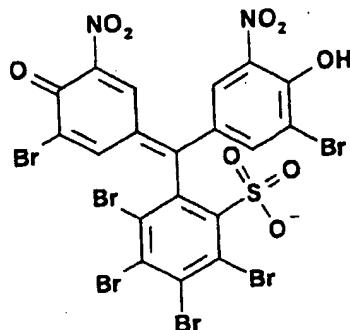
environment than when free in solution, and effectively develop a lower pKa for the ionization essential to color change. If the dye, in the absence of a protein, also responds as a pH sensitive color indicator within the selected target pH range, then added proteins will generally not be detected accurately.

5 The dyes to be used in the present invention generally provide a detectable response in the presence of total protein from about 2 to about 500 mg/dL protein. The dyes preferably provide a detectable and particularly useful response to total protein from about 15 to about 300 mg/dL protein. It is contemplated that the amount of dye may be varied to give a detectable and useful response outside of such a preferred range.

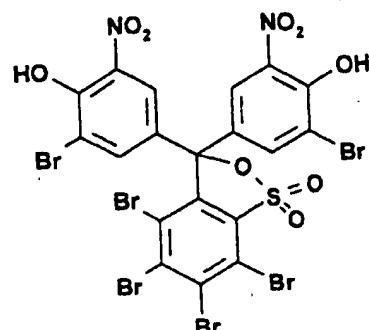
10 Examples of these dyes include substituted phenolsulfonephthaleins, pyrogallol red or combinations thereof. It is contemplated that other dyes that tend to have a detectable indication in the dye absorbance or fluorescence spectrum (as indicating transition) in the pH range of from about 2 to about 3 in the absence of protein and a different indication, under otherwise the same conditions, but in the presence of protein
15 may be used in the present invention. The pH for giving a different detectable response between the presence and absence of protein, should be within the pH range maintainable by the buffer being used in the present invention.

20 For example, it is contemplated that dyes such as bromochlorophenol blue (3',3"-dibromo-5',5"-dichlorophenolsulfonephthalein); bromophenol blue (tetrabromophenol blue); basic fuchsin (basic red 9); basic violet 14; martius yellow (acid yellow 24); phloxine B (acid red 92); methyl yellow (solvent yellow 2); congo red (direct red 28); methyl orange (acid orange 52); and ethyl orange (4-(4-diethylaminophenylazo)benzenesulfonic acid) may be useful in the present invention.
25 Such dyes may be used in combination with dyes such as phenolsulfonephthaleins and pyrogallol red.

The phenolsulfonephthalein indicators particularly useful in the present invention are represented by the following structures, H and I:



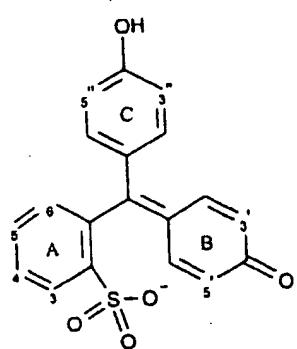
Structure H



Structure I

Structure H represents the structure of a phenolsulfonephthalein derivative in protic solvents such as water or an alcohol, while Structure I represents the form that predominates in the dry state or in aprotic solvents such as ethers and acetonitrile. The phenolsulfonephthalein "protein error" indicators are pH indicators that include an ionizable proton having a pKa value such that at particular pH's the proton is not ionized unless in the presence of protein.

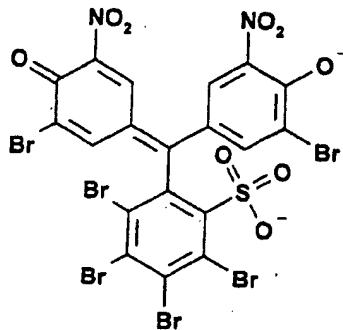
The pKa value of a general phenolsulfonephthalein indicator shown below, Structure J, is the pH at which one half of the number of indicator molecules include the deprotonated C ring phenolic hydroxyl group. In the case of the phenolsulfonephthalein protein error indicators illustrated above, two deprotonation events occur to cause an observable color change. The first deprotonation removes the proton from the aryl sulfonic acid on the A ring to yield the ion illustrated below as Structure J.



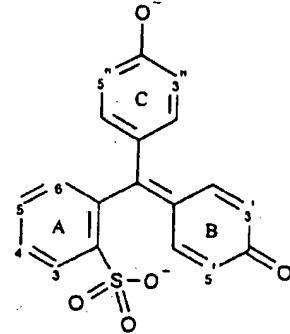
Structure J

The pKa of this proton is less than one, resulting in the ionization of this moiety at all useful pH values. This ionizable group is also responsible for the aqueous solubility of these compounds. The second deprotonation involves the release of a proton from the C ring phenolic hydroxyl to yield the dianion as shown in Structures K and L below.

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Structure K



Structure L

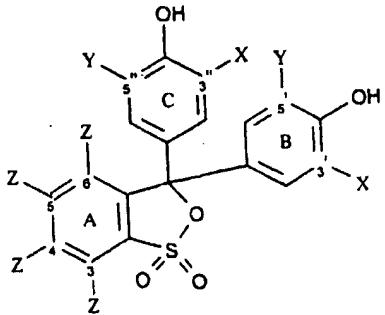
With this type of protein error indicator, the second deprotonation causes the observable color change that is indicative of the presence of protein in the sample being tested. The phenolsulfonephthalein protein error indicator is typically applied to an absorbent matrix material along with a buffer to provide an environment of constant pH, so that one can rely on the color change being the result of the presence of protein rather than the result of a pH change upon contact with the test fluid.

Only those phenolsulfonephthalein protein error indicators which have a second pKa allowing the second deprotonation, in the presence of protein, to take place at a pH within a range of from about 2.0 to about 3.0 are useful in the present invention. Those phenolsulfonephthalein protein error indicators that have a second pKa allowing the second deprotonation, in the absence of protein, to occur outside of the maintained pH may be useful in the invention. Preferably, the second pKa, in the absence of protein, takes place at a pH not within the range of from about 2.4 to about 3.0 and, most preferably, at a pH above 3.0. It is contemplated that an ionization sensitive to protein presence that also gives a detectable indication (e.g., a color response) may be a first, second, etc. deprotonation with a different assortment of ionizable substituents.

Many phenolsulfonephthalein protein error indicators have their second pKa in the appropriate range and are, therefore, suitable for use in the present invention. Useful

phenolsulfonephthaleins include those that are substituted with electron withdrawing and/or electron donating groups such as amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen, nitro, and cyano on the A, B or C rings of Structures J and L. It is contemplated that one or more of the rings may still contain a hydrogen. The particular substituent(s) and their positions on the dye molecule are not critical so long as the resulting dye exhibits a pKa in the appropriate range, and retains desirable physical properties, such as solubility, protein affinity and color. Unsubstituted phenolsulfonephthaleins are not suitable because their pKa's are not in the appropriate range. It has been discovered that the octa substituted sulfonephthalein indicators, i.e., those phenolsulfonephthalein derivatives that are substituted at the 3', 3'', 5', 5'', 3, 4, 5 and 6 positions, are particularly suitable for use in the present invention. These indicators are preferably substituted with halogen and nitro groups and may be represented by

Structure M:



Structure M

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where: X is nitro and Y and Z are chlorine, bromine or iodine. Also useful are the nitro substituted polyhalogenated phenolsulfonephthaleins disclosed in U.S. Pat. No. 5,279,790 wherein Y is nitro and X and Z are chlorine, bromine or iodine. These indicators are said to have the ability to detect from about 2 to 500 mg/dL of protein in a fluid test sample. However, use of the assay system of the present invention would be desirable in situations in which proteinuria greater than 30 mg/dL is measured and microalbuminuria at 2 mg/dL is not measured.

For example, in the case of diagnosing children for renal disease, it is not desirable to measure microalbumin as this response may give a false indication of disease due to its variable nature. It is evident that the aromatic rings of the phenolsulfonephthalein

indicators useful in the present invention can bear a variety of substituent groups. Such substituent groups are limited only by the ability of one of ordinary skill in the art to prepare stable compounds that have the appropriate protein error indicator properties to render them suitable for use in the present invention.

Some specific examples of the phenolsulfonephthalein dyes are those substituted at the 5', 5" positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or iodide. For example, phenolsulfonephthalein dyes that may be used include 5, 5"-Dinitro-3', 3", 3, 4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6,-hexabromophenolsulfonephthalein.

As discussed above, a dye that may be used is pyrogallol red. Pyrogallol red typically includes the use of a molybdate or tungstate salt that assists in providing rich color. U.S. Patent No. 5,399,498 discloses that tungstate reacts with the indicator to form a complex whose color shifts in the presence of protein in a manner similar to that of molybdenum. U.S. Patent No. 5,399,498 also discloses the use of phytic acid or derivatives thereof to reduce background interference in this sort of assay. While this assay is very good at detecting low levels of protein in a fluid test sample such as urine, its resolution drops off dramatically at higher protein concentrations, particularly at protein concentrations greater than about 150 mg/dL. This method is a total protein assay since the response is not dependent on the type of protein present in the test sample. Thus, human serum albumin (HSA) at 15 mg/dL provides the same response as IgG at 15 mg/dL. Since total protein assays are useful in the detection of certain disease states, and the higher the total protein spillover into urine the more serious the potential problem, a total protein assay that provides a detectable response at high protein concentrations is desirable. The molybdate/tungstate assay, however, is not effective for the detection of high levels of protein, i.e. above about 150 mg/dL, because of the strong affinity of the dye for protein.

Other Possible Components

The assay device or component may contain any number of surfactants, detergents, background dyes, enhancer polymers or chelating agents that are known in the art for use with protein detection methods based on dye binding. These assay devices or components typically include a protic solvent such as a water/methanol mixture in the

concentrations set out in Table 1. Typically, the solution will contain a chelating agent such as phytic acid and/or oxalic acid to inhibit or prevent interference from other components in the urine test sample. The acids in the paper dip process may be adjusted to a preferred pH of from about 2.4 to about 3 by adding sodium hydroxide or the like.

TABLE 1

		<u>Optional Components</u>	
<u>Ingredient</u>	<u>Function</u>	<u>Conc. Used</u>	<u>General Range</u>
Water	Solvent	95 mL	--
Methanol	Solvent	5 mL	0-40 mL
Phytic Acid	Chelator	1 g (15 mM)	0-500 mM
Oxalic Acid	Chelator	0.11 g (12 mM)	0-40 mM

EXAMPLES

One example of forming a dry device, such as a diagnostic strip, includes dipping a material such as paper into an aqueous solution. The diagnostic strip may be made from various materials, but is typically made from paper attached to a plastic carrier. One example of a suitable paper is Ahlstrom 204. It is contemplated that these strips may be made from other materials such as natural or synthetic woven, or nonwoven materials.

According to one embodiment, a first aqueous solution is formed by the addition of three submixes (Submixes 1-3). To form Submix 1, 10 ml of methanol, 1.76 ml of 125 mM TRIS ((tris(hydroxymethyl)aminomethane)/methanol) and 44.0 mg of pyrogallol red were added together. These components were mixed from about 20 to about 30 minutes to form Submix 1. To form Submix 2, 4.10 g (3.20 mL) of 40% phytic acid, 8.00 mL of 5% aqueous PVA (polyvinyl alcohol) of 31-50K, 11.2 ml of 1N NaOH and 0.654 mL of 100 mg/mL of sodium molybdate, and 45.6 mL of water were added together. The NaOH was added to adjust the pH to approximately 2.3. These components were mixed for about 60 minutes to form Submix 2. To form Submix 3, 217.5 mg of disodium oxalate, 120.0 mL of 500mM L-citrulline with a pH of 2.5, and 100.0 mg of Cibacron Brilliant Yellow were added together. The components were mixed for about 30 minutes to form Submix 3.

Submix 1 and Submix 2 were mixed together for about 10 minutes and then Submix 3 was added. The pH was 2.6 and the combined Submixes 1-3 was stabilized for 4 hours and adjusted to volume. Submixes 1-3 formed a First Dip solution of about 200

mL. The paper was dipped into the First Dip solution and dried in a three-stage oven at 50/50/70°C with a 1 inch air flow. The paper being used was Ahlstrom 204 paper.

The paper was then dipped into a Second Dip solution of 160 mL. The Second Dip solution was made by mixing 144 mL of toluene, 16 mL of THF (stabilized), 0.48 g of KOK (a polypropyleneglycol carbonate) such as disclosed in U.S. Patent No. 5,424,215 and 36.57 mg of DNHB (5, 5" Dinitro - 3', 3", 3, 4, 5, 6-hexabromophenolsulfonephthalein). The paper was dried in a three-stage oven at 50°/50°/50°C with a 1 inch air flow to form the active pad of the diagnostic strips. The diagnostic strips include the active pad that is adhered to a polymeric strip.

10 While particular embodiments and applications of the present invention have been illustrated and described, it is to be understood that the invention is not limited to the precise construction and compositions disclosed herein and that various modifications, changes, and variations may be apparent from the foregoing descriptions without departing from the spirit and scope of the invention as defined in the appended claims.

WHAT IS CLAIMED IS:

2 1. In an assay for the determination of protein in an aqueous test fluid which
comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
4 cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or
combinations thereof, the buffer is added in sufficient quantity to maintain the pH of the
6 assay including the test fluid at a selected target pH range within a range of from about
2.0 to about 3.0 and (b) a dye having a pKa which enables it to operate as a protein
8 indicator at the target pH range and whose affinity for protein is such that it will provide a
detectable response in the presence of greater than about 15 mg/dL protein to thereby
10 render the assay suitable for the detection of total protein in the test fluid.

12 2. The method of claim 1, wherein the buffer is selected from cyanoacetic
acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations
14 thereof.

16 3. The method of claim 1, wherein the buffer is added in a sufficient quantity
to maintain the pH of the assay including the test fluid within a range of from about 2.4 to
18 about 3.0.

20 4. The method of claim 1, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

22 5. The method of claim 4, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is octasubstituted with
24 amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen,
26 nitro, cyano, or combinations thereof.

28 6. The method of claim 4, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is substituted at the 5', 5"
30 positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or
iodide.

2 7. The method of claim 4, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is 5,5"-Dinitro-3', 3", 3,
4 4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6,-
hexabromophenolsulfonephthalein.

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8 8. The method of claim 1, wherein the dye is pyrogallol red and further
includes combining the test fluid with a molybdate or tungstate salt.

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9. The method of claim 1 wherein the buffer and dye are absorbed in a test
strip of absorbent material.

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14 10. The method of claim 1, wherein the detectable response provides good
resolution at protein concentrations of from about 15 to about 300 mg/dL.

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11. The method of claim 1, wherein the dye operates as a color indicator in the
presence of protein at a pH within the range of from about 2.0 to about 3.0.

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20 12. The method of claim 11, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.4 to about 3.0.

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13. The method of claim 1, wherein the buffer maintains the pH of the assay
including the test fluid at a target pH plus or minus about 0.2 pH units.

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26 14. In an assay for the determination of protein in an aqueous test fluid which
comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or
28 combinations thereof, the buffer is added in sufficient quantity to maintain the pH of the
assay including the test fluid within about +/- 0.2 pH units of a selected pH range within a
30 range of from about 2.0 to about 3.0 and (b) a dye having a pKa which enables it to
operate as a color indicator in the presence of protein at the target pH range and whose

affinity for protein is such that it will provide a detectable response in the presence of
2 greater than about 2 to about 500 mg/dL protein to thereby render the assay suitable for
the detection of total protein in the test fluid.

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15. A dry device for use in determining protein levels in a fluid test sample
which comprises an absorbent material having absorbed therein (a) a buffer selected from
a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl
phosphonic acid, sarcosine, saccharin, or combinations thereof, the buffer is added in
sufficient quantity to maintain the pH of the assay including the test fluid at a selected
target pH range within a range of from about 2.0 to about 3.0 and (b) a dye having a pKa
which enables it to operate as a protein indicator at the target pH range and whose affinity
for protein is such that it will provide a detectable response in the presence of greater than
15 mg/dL protein upon contact between the device and the fluid test sample to thereby
render the device suitable for the detection of total protein in the fluid test sample.

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19. The dry device of claim 15, wherein the buffer is selected from cyanoacetic
acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations
thereof.

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17. The dry device of claim 15, wherein the buffer is added in a sufficient
quantity to maintain the pH of the assay including the test fluid within a range of from
about 2.4 to about 3.0.

24
18. The dry device of claim 15, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

26
28
30
19. The dry device of claim 18, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is octasubstituted with
amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen,
nitro, cyano, or combinations thereof.

20. The dry device of claim 18, wherein the dye is a substituted
2 phenolsulfonephthalein, the substituted phenolsulfonephthalein is substituted at the 5', 5"
positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or
4 iodide.

6 21. The dry device of claim 18, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is 5,5"- Dinitro-3', 3", 3,
8 4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6,-
hexabromophenolsulfonephthalein.
10

12 22. The dry device of claim 15, wherein the dye is pyrogallol red and further
includes combining the test fluid with a molybdate or tungstate salt.

14 23. The dry device of claim 15, wherein the detectable response provides good
resolution at protein concentrations of from about 15 to about 300 mg/dL.
16

18 24. The method of claim 15, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.0 to about 3.0.
20

22 25. The method of claim 24, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.4 to about 3.0.
26

24 26. The method of claim 15, wherein the buffer maintains the pH of the assay
including the test fluid at a target pH plus or minus about 0.2 pH units.
28

26 27. A dry device for use in determining protein levels in a fluid test sample
which comprises an absorbent material having absorbed therein (a) a buffer selected from
28 a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl
phosphonic acid, sarcosine, saccharin, or combinations thereof, the buffer is added in
30 sufficient quantity to maintain the pH of the assay including the test fluid within about +/-
0.2 pH units of a selected target pH range within a range of from about 2.0 to about 3.0

and (b) a dye having a pKa which enables it to operate as a color indicator in the presence
2 of protein at the selected target pH range and whose affinity for protein is such that it will
provide a detectable response in the presence of from about 2 to about 500 mg/dL protein
4 upon contact between the device and the fluid test sample to thereby render the device
suitable for the detection of total protein in the fluid test sample.

6
28. In an assay for the determination of protein in an aqueous test fluid which
8 comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
or combinations thereof, the buffer is added in sufficient quantity to maintain the pH of
10 the assay including the test fluid at a selected pH range within a range of from about 2.0
to about 3.0 and (b) a dye having a pKa which enables it to operate as a protein indicator
12 at the target pH range and whose affinity for protein is such that it will provide a
detectable response in the presence of greater than about 15 mg/dL protein to thereby
14 render the assay suitable for the detection of total protein in the test fluid.

16 29. The method of claim 28, wherein the buffer is citrulline.

18 30. The method of claim 28, wherein the buffer is malonic acid.

20 31. The method of claim 28, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

22 32. The method of claim 28, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.0 to about 3.0.
24

26 33. The method of claim 32, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.4 to about 3.0.
28

30 34. A dry device for use in determining protein levels in a fluid test sample
which comprises an absorbent material having absorbed therein (a) a buffer selected from
a buffer selected from citrulline, malonic acid or combinations thereof, the buffer is added

in sufficient quantity to maintain the pH of the assay including the test fluid at a selected
2 target pH range within a range of from about 2.0 to about 3.0 and (b) a dye having a pKa
which enables it to operate as a protein indicator at the target pH range and whose affinity
4 for protein is such that it will provide a detectable response in the presence of greater than
15 mg/dL protein upon contact between the device and the fluid test sample to thereby
6 render the device suitable for the detection of total protein in the fluid test sample.

8 35. The dry device of claim 34, wherein the buffer is citrulline.

10 36. The dry device of claim 34, wherein the buffer is malonic acid.

12 37. The dry device of claim 34, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

14

16 38. The method of claim 34, wherein the dye operates as a color indicator at a
pH within the range of from about 2.0 to about 3.0 in the presence of protein.

18 39. The method of claim 38, wherein the dye operates as a color indicator at a
pH within the range of from about 2.4 to about 3.0 in the presence of protein.

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(54) Title: TOTAL PROTEIN DETECTION METHODS AND DEVICES AT LOW PH

(57) Abstract: An assay for the determination of protein in an aqueous test fluid that comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye also has affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein to thereby render the assay suitable for the detection of total protein in the test fluid. The buffer and dye may be absorbed in a test strip of absorbent material.

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B. FIELDS SEARCHED

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EPO-Internal, WPI Data, BIOSIS

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PCT/IB	02/02321

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- (71) Applicant (*for all designated States except US*): BAYER HEALTHCARE LLC [US/US]; 333 Coney Street, East Walpole, MA 02032 (US).
- (72) Inventors; and
- (75) Inventors/Applicants (*for US only*): PROFITT, James, A. [US/US]; 1805 Barclay Place, Goshen, IN 46528 (US). ORN, Alexander, H. [US/US]; 305 Bison Ridge, Nappanee, IN 46550 (US). FARR, Jennifer [US/US]; 2320 E. 600 N., Albion, IN 46701 (US).
- (74) Agent: LEVY, Elizabeth, A.; Bayer Corporation, 63 North Street, Medfield, MA 02052 (US).
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TOTAL PROTEIN DETECTION METHODS AND DEVICES AT LOW pH

5

FIELD OF THE INVENTION

The present invention relates generally to methods and devices of detecting proteins. Specifically, the invention is directed to methods and devices for detecting proteins at lower pH's using various buffers.

10

BACKGROUND OF THE INVENTION

Methods for the detection of urinary proteins are often more sensitive to albumin than to other urinary proteins. It is important, however, to detect proteins other than albumin, especially in the case of Bence-Jones proteinuria. The detection of proteins should cover a wide range of protein concentrations since the decision levels and recommended actions to be taken by clinicians will vary depending on the concentration of protein detected in a patient's urine. For example, persistent proteinuria greater than 50 mg/dL represents strong evidence of renal disease whereas a protein level of greater than 300 mg/dL is consistent with a diagnosis of nephrotic syndrome. A concentration of protein in urine of greater than 800 mg/dL suggests massive protein loss and warrants a renal biopsy and/or steroid therapy. Accordingly, it is apparent that a test for protein in urine should be effective over a wide range of protein values.

Various methods for the determination of protein in aqueous fluid have been reported in the literature. These methods include the Kjeldahl method, biuret method, Lowery method, dyestuff combination method, UV method and fluorometric method. In general, proteins react with various substance including dyes such as bromphenol blue, coomassie brilliant blue and eosine, as well as metal ions such as silver (I), copper (II), zinc (II) and lead (II).

Urine contains amounts of salt that vary between individuals. Some of these salts may act as a buffer when a diagnostic assay demands operation at a pH different than that of the urine sample. For example, urine sample with salts such as bicarbonate, acetate or phosphate tend to resist the lowering of pH. Creatinine, a common component of urine, may also act as a buffer in resisting the lowering of pH. The result of this buffering is that

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some urine samples will cause a strip pH to be higher than optimal. A higher than optimal strip pH may result in color generation of the protein indicator dyes, providing a false indication of the presence of protein. It is desired to have a buffering system that reduces such urine effects, while not interacting with the chemistry occurring.

5 Accordingly, a need exists for a method and device for detecting proteins over a wide range of clinical concentrations and pH's.

SUMMARY OF THE INVENTION

According to one embodiment, an assay for the determination of protein in an aqueous test fluid comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye has an affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein, to thereby render the assay suitable for the detection of total protein in the test fluid.

According to another embodiment, a dry device for use in determining protein levels in a fluid test sample comprises an absorbent material having absorbed therein a buffer and a dye. The buffer is selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range. The dye has an affinity for protein such that it will provide a detectable response in the presence of greater than 15 mg/dL protein upon contact between the device and the fluid test sample, to thereby render the device suitable for the detection of total protein in the fluid test sample.

According to a further embodiment, an assay for the determination of protein in an aqueous test fluid comprises combining the test fluid with a buffer and a dye. The buffer is selected from citrulline, malonic acid, or combinations thereof. The buffer is added in

sufficient quantity to maintain the pH of the assay including the test fluid at a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range and which has an affinity for protein such that it will provide a detectable response in the presence of greater than about 15 mg/dL protein, to thereby render the assay suitable for the detection of total protein in the test fluid.

According to yet a further embodiment, a dry device for use in determining protein levels in a fluid test sample comprises an absorbent material having absorbed therein a buffer and a dye. The buffer is selected from citrulline, malonic acid or combinations thereof. The buffer is added in sufficient quantity to maintain the pH of the assay including the test fluid within a selected target pH range within a range of from about 2.0 to about 3.0. The dye has a pKa which enables it to operate as a protein indicator at the target pH range and which has affinity for protein such that it will provide a detectable response in the presence of greater than 15 mg/dL protein upon contact between the device and the fluid test sample, to thereby render the device suitable for the detection of total protein in the fluid test sample.

DETAILED DESCRIPTION OF THE ILLUSTRATED EMBODIMENT

The assay for protein of the present invention can be carried out in either the wet or dry format. It is most conveniently carried out in the form of an absorbent test strip impregnated with (a) a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof, and (b) a dye having a pKa which enables it to operate as a protein indicator (e.g., a color indicator) at a pH of from about 2.0 to about 3.0.

Buffers

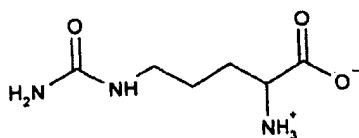
The buffer to be used in the present invention is added in sufficient quantity to maintain the pH of the assay including the test fluid within a range of from about 2.0 to about 3.0. The buffer may have a prior adjustment of pH that is commonly accomplished by adding acid or base to a solution of the buffer chemical to achieve a particular pH, or by adding a mixture of the buffer chemical and its salt in selected proportions to give a particular pH. The buffer is preferably added to maintain the pH of the assay including the test fluid within about +/- 0.2 pH units of the selected target pH. The buffer is

preferably added in sufficient quantity to maintain the pH of the assay including the test fluid within a range of from about 2.4 to about 3.0.

The preferred buffers are citrulline, malonic acid or combinations thereof. Buffers may be selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations thereof. Various concentrations of these buffers may be used, but the concentrations of the buffers are generally from about 200 to about 500 mM.

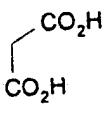
Citrulline may be in the form of L-citrulline, D-citrulline and D,L-citrulline and is represented by structure A.

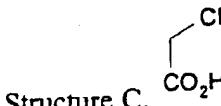
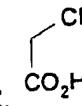
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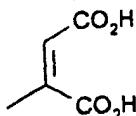


Structure A

Malonic acid that is useful in the present invention is represented by Structure B,

15  Cyanoacetic acid that is useful in the present invention is represented by

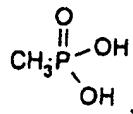
 Structure C,  Citraconic acid that is useful in the present invention is represented by Structure D.



Structure D

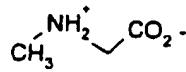
20

Methyl phosphonic acid that is useful in the present invention is represented by Structure E.



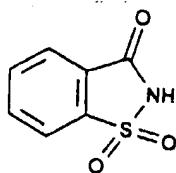
Structure E

Sarcosine that is useful in the present invention is represented by Structure F.



Structure F

Saccharin that is useful in the present invention is represented by Structure G.



10

Structure G

It is believed that citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine and/or saccharin can be obtained commercially, such as from a company like Aldrich Chemical Company.

15

Dyes

The dyes to be used in the present invention have pKa's that enable them to operate as protein indicators at a pH within a range of from about 2.0 to about 3.0. For example, the dyes may have pKa's that indicate a color difference when protein is present at a selected target pH range within the pH range of from about 2.0 to about 3.0. The selected target pH range is preferably within about +/- 0.2 pH units of the selected target pH. Additionally, the dyes may have pKa's that do not have a significant color difference in the absence of protein within the selected target pH range. The dyes to be used in the present invention preferably have pKa's that enable them to operate as protein indicators at a pH within a range of from about 2.4 to about 3.0. While not being bound by theory, it is believed that such dyes, when bound to a protein, are exposed to a different

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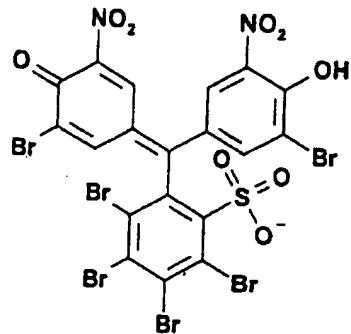
environment than when free in solution, and effectively develop a lower pKa for the ionization essential to color change. If the dye, in the absence of a protein, also responds as a pH sensitive color indicator within the selected target pH range, then added proteins will generally not be detected accurately.

5 The dyes to be used in the present invention generally provide a detectable response in the presence of total protein from about 2 to about 500 mg/dL protein. The dyes preferably provide a detectable and particularly useful response to total protein from about 15 to about 300 mg/dL protein. It is contemplated that the amount of dye may be varied to give a detectable and useful response outside of such a preferred range.

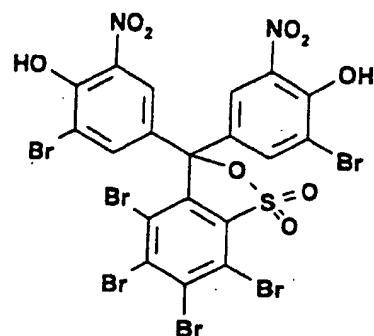
10 Examples of these dyes include substituted phenolsulfonephthaleins, pyrogallol red or combinations thereof. It is contemplated that other dyes that tend to have a detectable indication in the dye absorbance or fluorescence spectrum (as indicating transition) in the pH range of from about 2 to about 3 in the absence of protein and a different indication, under otherwise the same conditions, but in the presence of protein
15 may be used in the present invention. The pH for giving a different detectable response between the presence and absence of protein, should be within the pH range maintainable by the buffer being used in the present invention.

For example, it is contemplated that dyes such as bromochlorophenol blue (3',3"-dibromo-5',5"-dichlorophenolsulfonephthalein); bromophenol blue (tetrabromophenol
20 blue); basic fuchsin (basic red 9); basic violet 14; martius yellow (acid yellow 24); phloxine B (acid red 92); methyl yellow (solvent yellow 2); congo red (direct red 28); methyl orange (acid orange 52); and ethyl orange (4-(4-diethylaminophenylazo)benzenesulfonic acid) may be useful in the present invention.
25 Such dyes may be used in combination with dyes such as phenolsulfonephthaleins and pyrogallol red.

The phenolsulfonephthalein indicators particularly useful in the present invention are represented by the following structures, H and I:



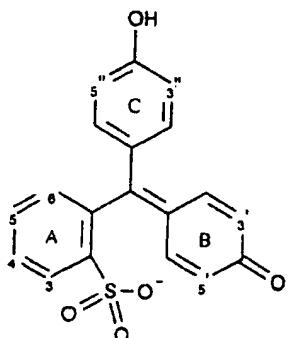
Structure H



Structure I

Structure H represents the structure of a phenolsulfonephthalein derivative in protic solvents such as water or an alcohol, while Structure I represents the form that predominates in the dry state or in aprotic solvents such as ethers and acetonitrile. The phenolsulfonephthalein "protein error" indicators are pH indicators that include an ionizable proton having a pKa value such that at particular pH's the proton is not ionized unless in the presence of protein.

The pKa value of a general phenolsulfonephthalein indicator shown below, Structure J, is the pH at which one half of the number of indicator molecules include the deprotonated C ring phenolic hydroxyl group. In the case of the phenolsulfonephthalein protein error indicators illustrated above, two deprotonation events occur to cause an observable color change. The first deprotonation removes the proton from the aryl sulfonic acid on the A ring to yield the ion illustrated below as Structure J.

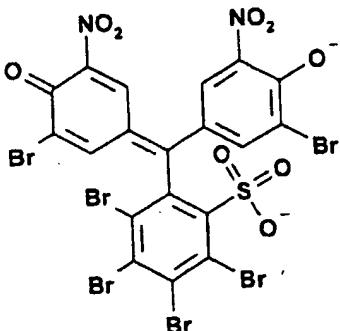


Structure J

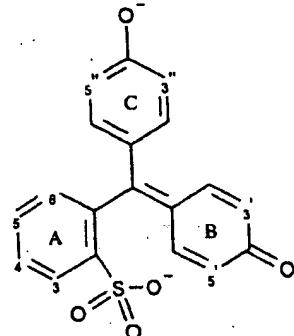
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The pKa of this proton is less than one, resulting in the ionization of this moiety at all useful pH values. This ionizable group is also responsible for the aqueous solubility of these compounds. The second deprotonation involves the release of a proton from the C ring phenolic hydroxyl to yield the dianion as shown in Structures K and L below.

5



Structure K



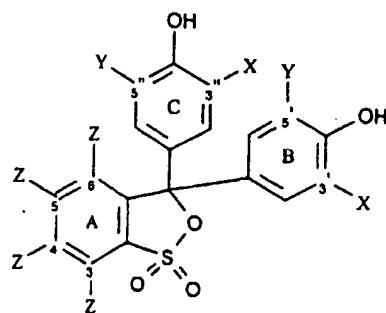
Structure L

With this type of protein error indicator, the second deprotonation causes the observable color change that is indicative of the presence of protein in the sample being tested. The phenolsulfonephthalein protein error indicator is typically applied to an absorbent matrix material along with a buffer to provide an environment of constant pH, so that one can rely on the color change being the result of the presence of protein rather than the result of a pH change upon contact with the test fluid.

Only those phenolsulfonephthalein protein error indicators which have a second pKa allowing the second deprotonation, in the presence of protein, to take place at a pH within a range of from about 2.0 to about 3.0 are useful in the present invention. Those phenolsulfonephthalein protein error indicators that have a second pKa allowing the second deprotonation, in the absence of protein, to occur outside of the maintained pH may be useful in the invention. Preferably, the second pKa, in the absence of protein, takes place at a pH not within the range of from about 2.4 to about 3.0 and, most preferably, at a pH above 3.0. It is contemplated that an ionization sensitive to protein presence that also gives a detectable indication (e.g., a color response) may be a first, second, etc. deprotonation with a different assortment of ionizable substituents.

Many phenolsulfonephthalein protein error indicators have their second pKa in the appropriate range and are, therefore, suitable for use in the present invention. Useful

phenolsulfonephthaleins include those that are substituted with electron withdrawing and/or electron donating groups such as amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen, nitro, and cyano on the A, B or C rings of Structures J and L. It is contemplated that one or more of the rings may still contain a hydrogen. The particular substituent(s) and their positions on the dye molecule are not critical so long as the resulting dye exhibits a pKa in the appropriate range, and retains desirable physical properties, such as solubility, protein affinity and color. Unsubstituted phenolsulfonephthaleins are not suitable because their pKa's are not in the appropriate range. It has been discovered that the octa substituted sulfonephthalein indicators, i.e., those phenolsulfonephthalein derivatives that are substituted at the 3', 3'', 5', 5'', 3, 4, 5 and 6 positions, are particularly suitable for use in the present invention. These indicators are preferably substituted with halogen and nitro groups and may be represented by Structure M:



15

Structure M

where: X is nitro and Y and Z are chlorine, bromine or iodine. Also useful are the nitro substituted polyhalogenated phenolsulfonephthaleins disclosed in U.S. Pat. No. 5,279,790 wherein Y is nitro and X and Z are chlorine, bromine or iodine. These indicators are said to have the ability to detect from about 2 to 500 mg/dL of protein in a fluid test sample. However, use of the assay system of the present invention would be desirable in situations in which proteinuria greater than 30 mg/dL is measured and microalbuminuria at 2 mg/dL is not measured.

For example, in the case of diagnosing children for renal disease, it is not desirable to measure microalbumin as this response may give a false indication of disease due to its variable nature. It is evident that the aromatic rings of the phenolsulfonephthalein

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indicators useful in the present invention can bear a variety of substituent groups. Such substituent groups are limited only by the ability of one of ordinary skill in the art to prepare stable compounds that have the appropriate protein error indicator properties to render them suitable for use in the present invention.

Some specific examples of the phenolsulfonephthalein dyes are those substituted at the 5', 5" positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or iodide. For example, phenolsulfonephthalein dyes that may be used include 5, 5"-Dinitro-3', 3", 3, 4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6-hexabromophenolsulfonephthalein.

As discussed above, a dye that may be used is pyrogallol red. Pyrogallol red typically includes the use of a molybdate or tungstate salt that assists in providing rich color. U.S. Patent No. 5,399,498 discloses that tungstate reacts with the indicator to form a complex whose color shifts in the presence of protein in a manner similar to that of molybdenum. U.S. Patent No. 5,399,498 also discloses the use of phytic acid or derivatives thereof to reduce background interference in this sort of assay. While this assay is very good at detecting low levels of protein in a fluid test sample such as urine, its resolution drops off dramatically at higher protein concentrations, particularly at protein concentrations greater than about 150 mg/dL. This method is a total protein assay since the response is not dependent on the type of protein present in the test sample. Thus, human serum albumin (HSA) at 15 mg/dL provides the same response as IgG at 15 mg/dL. Since total protein assays are useful in the detection of certain disease states, and the higher the total protein spillover into urine the more serious the potential problem, a total protein assay that provides a detectable response at high protein concentrations is desirable. The molybdate/tungstate assay, however, is not effective for the detection of high levels of protein, *i.e.* above about 150 mg/dL, because of the strong affinity of the dye for protein.

Other Possible Components

The assay device or component may contain any number of surfactants, detergents, background dyes, enhancer polymers or chelating agents that are known in the art for use with protein detection methods based on dye binding. These assay devices or components typically include a protic solvent such as a water/methanol mixture in the

concentrations set out in Table 1. Typically, the solution will contain a chelating agent such a phytic acid and/or oxalic acid to inhibit or prevent interference from other components in the urine test sample. The acids in the paper dip process may be adjusted to a preferred pH of from about 2.4 to about 3 by adding sodium hydroxide or the like.

5

TABLE 1

<u>Ingredient</u>	<u>Function</u>	<u>Optional Components</u>	<u>Conc. Used</u>	<u>General Range</u>
Water	Solvent		95 mL	--
Methanol	Solvent		5 mL	0-40 mL
Phytic Acid	Chelator		1 g (15 mM)	0-500 mM
Oxalic Acid	Chelator		0.11 g (12 mM)	0-40 mM

EXAMPLES

One example of forming a dry device, such as a diagnostic strip, includes dipping a material such as paper into an aqueous solution. The diagnostic strip may be made from various materials, but is typically made from paper attached to a plastic carrier. One example of a suitable paper is Ahlstrom 204. It is contemplated that these strips may be made from other materials such as natural or synthetic woven, or nonwoven materials.

According to one embodiment, a first aqueous solution is formed by the addition of three submixes (Submixes 1-3). To form Submix 1, 10 ml of methanol, 1.76 ml of 125 mM TRIS ((tris(hydroxymethyl)aminomethane)/methanol) and 44.0 mg of pyrogallol red were added together. These components were mixed from about 20 to about 30 minutes to form Submix 1. To form Submix 2, 4.10 g (3.20 mL) of 40% phytic acid, 8.00 mL of 5% aqueous PVA (polyvinyl alcohol) of 31-50K, 11.2 ml of 1N NaOH and 0.654 mL of 100 mg/mL of sodium molybdate, and 45.6 mL of water were added together. The NaOH was added to adjust the pH to approximately 2.3. These components were mixed for about 60 minutes to form Submix 2. To form Submix 3, 217.5 mg of disodium oxalate, 120.0 mL of 500mM L-citrulline with a pH of 2.5, and 100.0 mg of Cibacron Brilliant Yellow were added together. The components were mixed for about 30 minutes to form Submix 3.

Submix 1 and Submix 2 were mixed together for about 10 minutes and then Submix 3 was added. The pH was 2.6 and the combined Submixes 1-3 was stabilized for 4 hours and adjusted to volume. Submixes 1-3 formed a First Dip solution of about 200

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mL. The paper was dipped into the First Dip solution and dried in a three-stage oven at 50/50/70°C with a 1 inch air flow. The paper being used was Ahlstrom 204 paper.

The paper was then dipped into a Second Dip solution of 160 mL. The Second Dip solution was made by mixing 144 mL of toluene, 16 mL of THF (stabilized), 0.48 g of KOK (a polypropyleneglycol carbonate) such as disclosed in U.S. Patent No. 5,424,215 and 36.57 mg of DNHB (5, 5" Dinitro - 3', 3", 3, 4, 5, 6-hexabromophenolsulfonephthalein). The paper was dried in a three-stage oven at 50°/50°/50°C with a 1 inch air flow to form the active pad of the diagnostic strips. The diagnostic strips include the active pad that is adhered to a polymeric strip.

10 While particular embodiments and applications of the present invention have been illustrated and described, it is to be understood that the invention is not limited to the precise construction and compositions disclosed herein and that various modifications, changes, and variations may be apparent from the foregoing descriptions without departing from the spirit and scope of the invention as defined in the appended claims.

15

WHAT IS CLAIMED IS:

- 2 1. In an assay for the determination of protein in an aqueous test fluid which
4 comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
6 cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or
8 combinations thereof, the buffer is added in sufficient quantity to maintain the pH of the
10 assay including the test fluid at a selected target pH range within a range of from about
2.0 to about 3.0 and (b) a dye having a pKa which enables it to operate as a protein
indicator at the target pH range and whose affinity for protein is such that it will provide a
detectable response in the presence of greater than about 15 mg/dL protein to thereby
render the assay suitable for the detection of total protein in the test fluid.

- 12 2. The method of claim 1, wherein the buffer is selected from cyanoacetic
14 acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations
thereof.

- 16 3. The method of claim 1, wherein the buffer is added in a sufficient quantity
18 to maintain the pH of the assay including the test fluid within a range of from about 2.4 to
about 3.0.

- 20 4. The method of claim 1, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

- 22 5. The method of claim 4, wherein the dye is a substituted
24 phenolsulfonephthalein, the substituted phenolsulfonephthalein is octasubstituted with
amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen,
26 nitro, cyano, or combinations thereof.

- 28 6. The method of claim 4, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is substituted at the 5', 5"
30 positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or
iodide.

2 7. The method of claim 4, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is 5,5"-Dinitro-3', 3", 3,
4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6,-
hexabromophenolsulfonephthalein.

6 8. The method of claim 1, wherein the dye is pyrogallol red and further
8 includes combining the test fluid with a molybdate or tungstate salt.

10 9. The method of claim 1 wherein the buffer and dye are absorbed in a test
strip of absorbent material.

12 10. The method of claim 1, wherein the detectable response provides good
14 resolution at protein concentrations of from about 15 to about 300 mg/dL.

16 11. The method of claim 1, wherein the dye operates as a color indicator in the
presence of protein at a pH within the range of from about 2.0 to about 3.0.

18 12. The method of claim 11, wherein the dye operates as a color indicator in
20 the presence of protein at a pH within the range of from about 2.4 to about 3.0.

22 13. The method of claim 1, wherein the buffer maintains the pH of the assay
including the test fluid at a target pH plus or minus about 0.2 pH units.

24 14. In an assay for the determination of protein in an aqueous test fluid which
26 comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
cyanoacetic acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or
28 combinations thereof, the buffer is added in sufficient quantity to maintain the pH of the
assay including the test fluid within about +/- 0.2 pH units of a selected pH range within a
30 range of from about 2.0 to about 3.0 and (b) a dye having a pKa which enables it to
operate as a color indicator in the presence of protein at the target pH range and whose

affinity for protein is such that it will provide a detectable response in the presence of
2 greater than about 2 to about 500 mg/dL protein to thereby render the assay suitable for
the detection of total protein in the test fluid.

4

15. A dry device for use in determining protein levels in a fluid test sample
6 which comprises an absorbent material having absorbed therein (a) a buffer selected from
a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl
8 phosphonic acid, sarcosine, saccharin, or combinations thereof, the buffer is added in
sufficient quantity to maintain the pH of the assay including the test fluid at a selected
10 target pH range within a range of from about 2.0 to about 3.0 and (b) a dye having a pKa
which enables it to operate as a protein indicator at the target pH range and whose affinity
12 for protein is such that it will provide a detectable response in the presence of greater than
15 mg/dL protein upon contact between the device and the fluid test sample to thereby
14 render the device suitable for the detection of total protein in the fluid test sample.

16

16. The dry device of claim 15, wherein the buffer is selected from cyanoacetic
acid, citraconic acid, methyl phosphonic acid, sarcosine, saccharin, or combinations
18 thereof.

20

17. The dry device of claim 15, wherein the buffer is added in a sufficient
quantity to maintain the pH of the assay including the test fluid within a range of from
22 about 2.4 to about 3.0.

24

18. The dry device of claim 15, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

26

19. The dry device of claim 18, wherein the dye is a substituted
28 phenolsulfonephthalein, the substituted phenolsulfonephthalein is octasubstituted with
amino, amido, aromatic, alkyl, hydroxyl, thio, carboxylic, alkoxy, ketoxy, acetyl, halogen,
30 nitro, cyano, or combinations thereof.

20. The dry device of claim 18, wherein the dye is a substituted
2 phenolsulfonephthalein, the substituted phenolsulfonephthalein is substituted at the 5', 5"
positions with nitro and at the 3', 3", 3, 4, 5 and 6 positions with chloride, bromide or
4 iodide.

6 21. The dry device of claim 18, wherein the dye is a substituted
phenolsulfonephthalein, the substituted phenolsulfonephthalein is 5,5"-Dinitro-3', 3", 3,
8 4, 5, 6-hexabromophenolsulfonephthalein or 5'-Nitro-5"-Iodo-3', 3", 3, 4, 5, 6,-
hexabromophenolsulfonephthalein.

10 22. The dry device of claim 15, wherein the dye is pyrogallol red and further
12 includes combining the test fluid with a molybdate or tungstate salt.

14 23. The dry device of claim 15, wherein the detectable response provides good
resolution at protein concentrations of from about 15 to about 300 mg/dL.

16 24. The method of claim 15, wherein the dye operates as a color indicator in
18 the presence of protein at a pH within the range of from about 2.0 to about 3.0.

20 25. The method of claim 24, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.4 to about 3.0.

22 26. The method of claim 15, wherein the buffer maintains the pH of the assay
24 including the test fluid at a target pH plus or minus about 0.2 pH units.

26 27. A dry device for use in determining protein levels in a fluid test sample
which comprises an absorbent material having absorbed therein (a) a buffer selected from
28 a buffer selected from citrulline, malonic acid, cyanoacetic acid, citraconic acid, methyl
phosphonic acid, sarcosine, saccharin, or combinations thereof, the buffer is added in
30 sufficient quantity to maintain the pH of the assay including the test fluid within about +/-
0.2 pH units of a selected target pH range within a range of from about 2.0 to about 3.0

and (b) a dye having a pKa which enables it to operate as a color indicator in the presence
2 of protein at the selected target pH range and whose affinity for protein is such that it will
provide a detectable response in the presence of from about 2 to about 500 mg/dL protein
4 upon contact between the device and the fluid test sample to thereby render the device
suitable for the detection of total protein in the fluid test sample.
6

28. In an assay for the determination of protein in an aqueous test fluid which
8 comprises combining the test fluid with (a) a buffer selected from citrulline, malonic acid,
or combinations thereof, the buffer is added in sufficient quantity to maintain the pH of
10 the assay including the test fluid at a selected pH range within a range of from about 2.0
to about 3.0 and (b) a dye having a pKa which enables it to operate as a protein indicator
12 at the target pH range and whose affinity for protein is such that it will provide a
detectable response in the presence of greater than about 15 mg/dL protein to thereby
14 render the assay suitable for the detection of total protein in the test fluid.

16 29. The method of claim 28, wherein the buffer is citrulline.

18 30. The method of claim 28, wherein the buffer is malonic acid.

20 31. The method of claim 28, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.
22

32. The method of claim 28, wherein the dye operates as a color indicator in
24 the presence of protein at a pH within the range of from about 2.0 to about 3.0.

26 33. The method of claim 32, wherein the dye operates as a color indicator in
the presence of protein at a pH within the range of from about 2.4 to about 3.0.
28

34. A dry device for use in determining protein levels in a fluid test sample
30 which comprises an absorbent material having absorbed therein (a) a buffer selected from
a buffer selected from citrulline, malonic acid or combinations thereof, the buffer is added

in sufficient quantity to maintain the pH of the assay including the test fluid at a selected
2 target pH range within a range of from about 2.0 to about 3.0 and (b) a dye having a pKa
which enables it to operate as a protein indicator at the target pH range and whose affinity
4 for protein is such that it will provide a detectable response in the presence of greater than
15 mg/dL protein upon contact between the device and the fluid test sample to thereby
6 render the device suitable for the detection of total protein in the fluid test sample.

8 35. The dry device of claim 34, wherein the buffer is citrulline.

10 36. The dry device of claim 34, wherein the buffer is malonic acid.

12 37. The dry device of claim 34, wherein the dye is a substituted
phenolsulfonephthalein, pyrogallol red or combinations thereof.

14 38. The method of claim 34, wherein the dye operates as a color indicator at a
16 pH within the range of from about 2.0 to about 3.0 in the presence of protein.

18 39. The method of claim 38, wherein the dye operates as a color indicator at a
pH within the range of from about 2.4 to about 3.0 in the presence of protein.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/IB 02/02321

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 G01N33/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 7 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>WATANABE N ET AL: "Urinary Protein as Measured with a Pyrogallol Red-Molybdate Complex, Manually and in a Hitachi 726 Automated Analyzer"</p> <p>CLINICAL CHEMISTRY, AMERICAN ASSOCIATION FOR CLINICAL CHEMISTRY. WINSTON, US, vol. 32, no. 8, 1986, pages 1551-1554, XP002091174</p> <p>ISSN: 0009-9147</p> <p>page 1551</p> <p>---</p>	1-39
Y	<p>EP 0 361 244 A (MILES, INC)</p> <p>4 April 1990 (1990-04-04)</p> <p>page 7, line 41 -page 13, line 15; claims 1-7</p> <p>---</p> <p>-/-</p>	1-39

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- *A* document defining the general state of the art which is not considered to be of particular relevance
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- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *Z* document member of the same patent family

Date of the actual completion of the international search

26 February 2003

Date of mailing of the international search report

06/03/2003

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INTERNATIONAL SEARCH REPORT

International Application No
PCT/IB U2/02321

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	US 5 424 215 A (MILES, INC) 13 June 1995 (1995-06-13) the whole document ----	1-39
Y	EP 0 785 429 A (BAYER CORPORATION) 23 July 1997 (1997-07-23) the whole document ----	1-39
Y	EP 0 907 081 A (BAYER CORPORATION) 10 September 1998 (1998-09-10) claims 1-10 -----	1-39

INTERNATIONAL SEARCH REPORT

Information on patent family members

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